Synthesis and Anticancer Molecular Docking Studies of Phenothiazine Derivatives

K. Venkatesan

Asst. Professor, CVR College of Engineering/ Humanities and Sciences Department, Hyderabad, India Email: venkippk@gmail.com

Abstract: Phenothiazine incorporated pyran derivatives are prepared by using N-alkyl-phenothiazine-3-carbaldehydes under ultrasonic/reflux condition by one step synthesis. Improvement in yield is observed when the reaction is carried out under ultrasonic condition compared to classical synthetic method. The synthesized compounds are characterized by using FT-IR, ¹H NMR, ¹³C NMR and mass spectral data. Anticancer docking studies are carried out for the synthesized compounds and different cancer target proteins namely lung cancer, colon cancer, breast cancer and pancreatic cancer and the results are reported.

Index Terms— Phenothiazine, Pyran, Anticancer docking and Ultrasonic Irradiation.

I. INTRODUCTION

Hetero aromatic compound containing Nitrogen and Sulfur atom in their backbone is being pursued as a thrust area of research in Chemistry due to its potential applications of these derivatives are used in pharma field. The combination of two or more pharmacophores in one molecule is an idea involved in designing of new biologically active chemical compound including natural alkaloids. Phenothiazine, a hetero aromatic tricyclic compound containing nitrogen and sulfur hetero atom in their ring has been well known for a period of hundred years. The first parent compound of unsubstituted phenothiazine was synthesized in 1883 by a synthetic chemist Bernthsen.

These derivatives were used in treatment of psychopathy for the first time and to get better results. Phenothiazine is a heterocyclic compound of possess lots of pharmaceutical applications such as antifungal [1], antibacterial [2-3], anti-inflammatory [4], anti-malarial [5], anti-psychotropic [6], antimicrobial [7], antitubercular [8-9] activities. It is reported that these derivatives have a few significant anticancer activities. These characteristics have generated a great interest in formulating and synthesizing new phenothiazine to investigate their anticancer activities. [10-11].

Phenothiazine derivatives also act as human cholinesterase inhibitors and on several occasions, these derivatives have been characterized as multidrug resistance (MDR) reversal agents [12-13]. The literature reports have shown that the anticancer activity of phenothiazine derivatives are determined by the substituents linked to Carbon-2 position of phenothiazine ring, extent of alkyl group connecting nitrogen atom attached at position (N-10) of the cyclic ring, also the terminal N-H group present in ring [14-15]. This activity is firmly bound to the nature of substituents attached

to phenothiazine ring rather than nature of side chain present [16].

Phenothiazine derivatives are used as neuroleptic drugs because they cross the blood brain barrier possess a strong affinity to lipid-rich tissues and lipid bilayers in neurons because of high degree of lipophilicity of phenothiazine derivatives [17]. In order to get active neuroleptic agents containing hydrogen atoms attached to nitrogen N-10 carbon-2 atoms are replaced by different groups attached at N-10 position such, as piperidin, aliphatic side chain and piperazine derivatives are given in literature [18]. They are also used as antipsychotic drugs because of their easily interaction with various receptors in central nervous system particularly which blocks the dopaminergic receptors [19].

A. Experimental procedure

General: All the chemicals were purchased from SD Fine Chemicals (India), Sisco Research Laboratory and Sigma-Aldrich (USA). Open capillary tubes were used to determine melting point of reported compound by a Buchi-530 melting point apparatus and the results were uncorrected. Perkin Elmer of FT-IR 1600 RX1 spectrophotometer used for recording FT-Infra Red spectra and potassium bromide used as discs. Proton and Carbon NMR were recorded using AV-400MHz and Bruker DPX 400 MHz spectrometer using deuterated chloroform and dimethyl sulphoxide a solvent by using an internal reference of Tetramethylsilane (TMS). Mass spectra were obtained using HR mass spectrometer and the compounds were dried under vacuum before analysis.

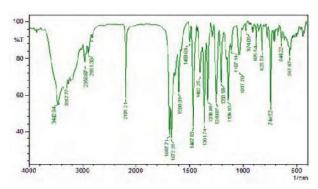
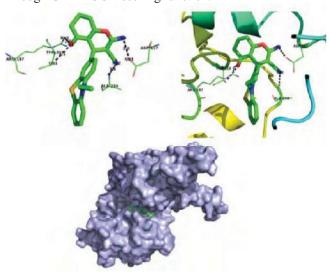


Figure 1. IR spectrum of compound 4a.

B. Docking Analysis

Geometry optimized structures for all new synthesized derivatives were obtained using iGEMDOCK automated docking program. The 3-dimensional coordinates of different target cancer proteins were selected from protein data bank (PDB). The PDB id of pancreatic cancer (1SVC), lung cancer (1MOX), colon cancer (4FLH), and breast cancer (2DSQ) were chosen for docking study. The 3D structure of ligand molecules and therapeutic target proteins were executed through GEMDOCK graphical interface. Before starting docking analysis, the output path was set. The default parameters were included such as population size (n=200), number of solutions (s=10) and generation (g=70) to compute the feasible binding mechanism of ligand moiety for each target protein. The docking run was initiated through GEMDOCK scoring function.



Figur. 2. Active sites of Rho-associated protein kinase 1 of compound **4d** by docking studies.

The binding pose of every ligand molecule was observed after docking analysis and their affinity with target proteins were analyzed. Binding geometries are predicted by visual examination which contributes essentially further development of a new compound. The best binding pose and total energy of each ligand molecules were analyzed using post docking analysis and the details were saved in output folder. Pymol automated docking software used to analyzed and visualized the protein-ligand binding site [20-21].

Protocol for the preparation of compound 2(a-b):

In a 50 mL two-necked RB flask a mixture of Phenothiazine (1.0 mmol), ethyl or methyl iodide (3.0 mmol) and DMF (25 mL) were taken. The solution was heated in an oil bath at 75°C, potassium tert-butoxide was added (1.5 mmol) and stirred for 24 hours. Completion of reaction was observed through thin layer chromatography, and cooled to room temperature and transferred into ice cooled water, the reaction mixture was extracted using chloroform and dried using sodium sulphate salt to get crude product. The impure products were subjected to column chromatography using hexane/ethyl ethanoate (4.5:0.5) to get pure compound as a colourless solid (yield: 81-83%).

Spectral data of compound 2a: mp. 96-98 °C; IR (KBr) vmax: 1130, 1280, 1327, 1384, 1440, 1483, 1570, 1591, 1788, 1899, 2826, 2937, 2981, 3053 3053, cm⁻¹; ¹H NMR (500 MHz, ppm, CDCl₃), $\delta_{\rm H}$ 3.40 (s, 3H, CH₃), 6.84 (d, 2H, J 8.0 Hz, C₃, C₆- Ph-H), 6.95-7.28 (m, 6H, Ph-H); ¹³C NMR

(125.787 MHz, ppm, CDCl₃), $\delta_{\rm C}$ 35.3, 114.0, 122.4, 123.4, 127.4, 145.8; Mass (EI): m/z [M⁺] calculated for C₁₃H₁₁NS: 212.0612; obtained: 212.2038.

Spectral data of compound 2b: mp. 102-104°C; IR (KBr) vmax: 1107, 1136, 1163, 1259, 1286, 1330, 1456, 1489, 1568, 2816, 2879, 2960, 3055 cm-1; 1H NMR (500 MHz, ppm, CDCl₃), $\delta_{\rm H}$ 1.46 (t, 3H, *J* 5.4Hz, CH₃), 3.96 (d, 2H, *J* 5.7Hz, CH₂), 6.90-7.28 (m, 8H, Ph-H); ¹³C NMR (125.787 MHz, ppm, CDCl₃), $\delta_{\rm C}$ 13.7, 41.6, 115.1, 122.3, 124.5, 127.2, 127.4, 145.0; Mass (EI): m/z [M⁺] calculated. for C₁₄H₁₃NS: 227.0769; obtained: 227.1685.

Protocol for the preparation of compound 3(a-b):

In a two necked RB flask a solution of POCl₃ (4.1 mmol) and freshly distilled dry DMF (4.7 mmol) was added drop wise at 0°C under inert or nitrogen atmospheric condition. The compound 2(a-b) (1.0 mmol) dissolved in 40 mL of DCM and the solution was added dropwise to the POCl₃/DMF complex at 30°C. The reaction mixture was stirred at 80°C in an oil bath for 16 hours. Completion of the reaction was monitored through thin layer chromatography and then cooled to room temperature and poured it to 200 grams of ice crystals. The reaction mixture was neutralized by adding sodium bicarbonate solution, then extracted with chloroform, dried using sodium sulphate salt and the excess solvent was evaporated using vacuum distillation to get crude product. The impure product was subjected to column chromatography hexane/ethyl ethanoate solvent; to get pure yellow coloured compound, yield obtained was 79-82%.

Spectral data of compound 3a: mp. 106-108 °C; IR (KBr) vmax: 1036, 1144, 1288, 1327, 1566, 1595, 1641, 1678, 2884, 3056 cm⁻¹; ¹H NMR (400 MHz, ppm, CDCl₃), $\delta_{\rm H}$ 3.37 (s, 3H, CH₃), 7.00-7.74 (m, 7H, Ph-H), 9.79 (s, 1H, CHO); ¹³C NMR (100.602 MHz, ppm, CDCl₃), $\delta_{\rm C}$ 35.6, 114.5, 115.4, 121.1, 122.3, 123.5, 126.9, 127.2, 128.0, 130.4, 130.8, 143.6, 150.4, 190.6; Mass (EI): m/z [M+] calculated. for C₁₄H₁₁NOS: 241.0351; obtained: 241.0187.

Spectral data of compound 3b: mp. 109-111 °C; IR (KBr) vmax: 1042, 1135, 1199, 1238, 1310, 1466, 1552, 1572, 1669, 2738, 2827, 2931, 2977, 3057 cm⁻¹; ¹H NMR (400 MHz, ppm, CDCl₃), $\delta_{\rm H}$ 1.45 (d, 3H, *J* 6.4Hz, CH₃), 3.97 (d, 2H, *J* 6.4Hz, CH₂), 7.62 (d, 1H, *J* 8.4Hz, C₇-Ph-H), 7.56 (s, 1H, C8- Ph-H), 6.89-7.26 (m, 5H, Ph-H), 9.78 (s, 1H, CHO); ¹³C NMR (100.602 MHz, ppm, CDCl₃), $\delta_{\rm C}$ 12.8, 42.4, 114.4, 115.6, 123.2, 124.4, 127.6, 128.2, 130.2, 143.0, 150.3, 190.0; Mass (EI): m/z [M⁺] calculated. for C₁₅H₁₃NOS: 255.1688; obtained: 255.1798.

Protocol for the preparation of compound 4(a-d).

Conventional method: In a 100 mL RB flask a mixture of compound 3a-b (1.0 mmol), 1,3-diketone (1.0 mmol), malononitrile 1.0 mmol, K_3PO_4 (15 mmol%) and 20% ethanol (5 ml) were refluxed with constant stirring at 80°C in an oil bath for the period of time listed in Table. 1 (completion of reaction monitored by thin layer chromatography using hexane/ethyl ethanoate as eluent). The reaction mixture on cooling to get precipitated crude

product and the solid obtained was filtered and recrystallization process was done by using ethanol solvent to get pure product.

Ultrasonic method: In a 25 mL beaker, a mixture of compound 3a-b (1.0 mmol), 1,3-diketones 1.0 mmol, malononitrile (1.0 mmol) in the presence of K_3PO_4 catalyst 15 mmol% and 20% ethanol (5 ml) were sonicated using ultrasonic probe at frequency range of 22 kHz for the given periods of time listed in Table. Completion of the reaction observed by thin layer chromatography, the precipitate obtained was filtered using funnel and washed with large amount of water and purified by recrystallization process using ethanol solvent.

Spectral data of compound 4a: mp. 194-196 °C; IR (KBr) vmax: 1037, 1103, 1203, 1249, 1330, 1361, 1402, 1467, 1598, 2189, 2891, 2958, 3257, 3442 cm⁻¹; ¹H NMR (400 MHz, ppm, CDCl₃), $\delta_{\rm H}$ 0.97 (s, 3H, CH₃), 1.03 (s, 3H, CH₃), 2.37 (q, 2H, *J* 7.8Hz, CH₂), 3.27 (s, 3H, N-CH₃), 4.25 (s, 1H, CH) 4.49 (s, 2H, NH₂), 6.67 (d, 1H, *J* 7.6Hz, C₄- Ph-H), 2.14 (q, 2H, *J* 6.6Hz, CH₂), 6.71 (d, 1H, *J* 7.6Hz, C₂-Ar-H), 6.84-7.20 (m, 5H, Ph-H); ¹³C NMR (100.602 MHz, ppm, CDCl₃), $\delta_{\rm C}$ 27.8, 28.6, 32.2, 34.6, 35.2, 40.4, 50.7, 63.2, 113.7, 118.5, 122.3., 123.0, 123.5, 125.8, 127.2, 127.4, 137.5, 144.8, 145.7, 157.4, 161.3, 195.8; Mass (EI): *m/z* [M[†]] calculated for C₂₅H₂₃N₃O₂S: 429.1511, obtained: 429.1514.

Spectral data of compound 4b: mp. 161-163 °C; IR (KBr) vmax: 1035, 1215, 1251, 1385, 1485, 1606, 1680, 2191, 2933, 2958, 3170, 3255, 3352 cm⁻¹; ¹H NMR (400 MHz, ppm, CDCl₃), $\delta_{\rm H}$ 0.99 (s, CH₃, 3H), 1.03 (s, CH₃, 3H), 1.32 (s, 3H, CH₃), 2.15 (s, 2H, CH₂), 2.37 (q, 2H, *J* 7.6Hz, CH₂), 3.82 (s, 2H, N-CH₂), 4.24 (s, 1H, CH), 4.45 (s, 2H, NH₂), 6.82-7.20 (m, 5H, Ph-H), 6.71 (d, 1H, *J* 7.3Hz, C₂-Ar-H), 6.75 (d, 1H, *J* 6.8Hz, C₂-Ar-H); ¹³C NMR (100.602 MHz, ppm, CDCl₃), $\delta_{\rm C}$ 12.9, 27.9, 28.6, 32.2, 34.5, 40.6, 41.7, 50.6, 63.4, 113.3, 14.8, 118.5, 122.1, 124.0, 126.1, 126.9, 127.1, 137.3, 144.1, 144.8, 157.3, 161.3, 195.8; Mass (EI): *m/z* [M⁺] calculated for C₂₆H₂₅N₃O₂S: 443.1667, obtained: 443.1663.

Spectral data of compound 4c: mp. 150-152 °C; IR (KBr) vmax: 1125, 1260, 1334, 1365, 1485, 1566, 1606, 1647, 1680, 2191, 2203, 2922, 3360 cm⁻¹; ¹H NMR (400 MHz, ppm, CDCl₃), δ_H 2.02-2.20 (m, CH₂, 2H), 2.28-2.35 (m, 2H, CH₂), 2.40-2.59 (m, CH₂, 2H), 3.32 (s, 3H, *N*-CH₃), 4.34 (s, 1H, CH), 4.52 (s, 2H, NH₂), 6.74 (d, 1H, *J* 7.2Hz, 4- Ph-H), 6.85 (d, 1H, *J* 7.5Hz, C₂- Ph-H), 6.91-7.23 (m, 6H, Ph-H); ¹³C NMR (100.602 MHz, ppm, CDCl₃), δ_C 20.0, 26.8, 35.9, 36.7, 401., 60.5, 114.0, 114.5, 121.9, 122.3, 124.1, 124.3, 125.3, 125.8, 127.3, 127.8, 129.0, 131.6, 143..2, 151.2, 157.4, 194.2; Mass (EI): *m/z* [M⁺] calculated for C₂₃H₁₉N₃O₂S: 401.1198, obtained: 401.1192.

Spectral data of compound 4d: mp. 143-145 °C; IR (KBr) vmax: 1120, 1271, 1365, 1483, 1576, 1600, 1648, 1679, 2190, 2922, 3268, 3382 cm⁻¹; ¹H NMR (400 MHz, ppm, CDCl₃), δ_H 1.25 (s, CH₃, 3H), 1.46-1.55 (m, CH₂, 2H), 2.02

(m, 2H, CH₂), 2.57 (q, 2H, J 6.6Hz, CH₂), 3.48 (s, 2H, N-CH₂), 4.33 (s, 1H, CH), 4.50 (s, 2H, NH₂), 6.74 (d, 1H, J 8.1Hz, C₄- Ph-H), 6.82 (d, 1H, J 8.1Hz, C₂- Ph-H), 6.93-7.25 (m, 5H, Ph-H); ¹³C NMR (100.602 MHz, ppm, CDCl₃), $\delta_{\rm C}$ 13.1, 20.2, 27.1, 36.9, 41.8, 42.8, 59.8, 114.5, 114.9, 115.3, 124.2, 127.3, 127.6, 127.9, 129.4, 131.6, 144.2, 150.4, 157.5, 194.4; Mass (EI): m/z [M⁺] calculated for C₂₄H₂₁N₃O₂S: 415.1534, obtained: 415.1530.

II. RESULTS AND DISCUSSION

Phenothiazine (1) was treated with methyl, ethyl iodide using potassium tertiary but-oxide catalyst and dry DMF used as a solvent and heated in an oil bath around 80 °C for 24 hours to yield alkyl substituted phenothiazine **2a-b** (Scheme 1). The compound **2a** showed a singlet peak at δ 3.40 ppm in ¹H NMR spectrum of corresponds to methyl group which is an evidence for the compound **2a** formation. The disintegration of N-H band at 3420 cm⁻¹ in IR spectra of **2a** supporting the formation of alkyl phenothiazine. The compound **2b** showed a quartet peak at δ 3.96 ppm and triplet peak at δ 1.47 ppm in ¹H NMR spectrum indicating the formation of CH₂ and CH₃ protons which is an evidence for compound **2b** formation. The compound **2b** showed peaks at δ 41.76 and 14.09 ppm in ¹³C NMR spectrum indicated the presence of CH₃ and CH₂ carbons in compound **2b**.

TABLE I
PHYSICAL DATA OF SYNTHESIZED COMPOUNDS

Compou	R	R_1	Reaction	Yield ^{a,}	m.p. (°C)
nd			Time ^a (min)	b(%)	
4a	CH ₃	CH ₃	165/7	85/87	195-197
4b	CH ₂ -CH ₃	CH ₃	170/8	83/86	160-162
4c	CH ₃	Н	175/8	84/86	152-155
4d	CH ₂ -CH ₃	Н	180/9	81/83	145-147

^aAt reflux temperature/ ultrasonic irradiation; ^bIsolated yields

The compound 2a-b on treatment with Vilsmeier Hack reaction to get compound 3a-b with a yield of 78-80%. The compound 3a and 3b showed a singlet peak at δ 9.76 and 9.78 ppm observed in ¹H NMR spectra indicating the formation of aldehyde hydrogen and the peaks at δ 190.68 and 190.10 ppm in ¹³C NMR spectra also confirming the presence of aldehyde carbon. The compound 3a and 3b showed the absorption peak at 1679 and 1665 cm⁻¹ in IR spectra indicated the presence of aldehyde carbonyl group which is also an evidence for the formation of 3a and 3b. The compound 4(a-d) was prepared by one step multi constituent reaction of dimedone, malono nitrile, 3(a-b) and K₃PO₄ catalyst were heated at 100 °C with equal ratio of Et-OH and water was used as a solvent for a given period of time listed in table and the same reaction was carried out using ultra sonication. The compound 4a showed absorption frequency at 3432 and 3263 cm⁻¹ in IR spectra due to amine group at 2187 cm⁻¹ due to CN and 1686 cm⁻¹ (Figure. 1) due to ketonic group which is also supporting the compound 4a formation.

SCHEME I SYNTHETIC PROTOCOL OF COMPOUND 4a-d.

The compound **4a** shows two singlet peaks at δ 1.02, 0.98 ppm corresponds to CH₃ hydrogens of cyclomethone, while CH₂ hydrogen of cyclomethone appeared as quartet peak δ 2.39, 2.13 and CH₃ hydrogen attached to nitrogen appeared at δ 3.29 as a singlet. The peak at δ 4.25 ppm belongs to benzylic methyl hydrogen and the singlet peak at δ 4.49 ppm belongs to -NH₂ hydrogen and which is clearly indicating the compound **4a** formation. The structures of all the synthesized derivatives were conformed through spectral techniques such as IR, ¹H, ¹³C NMR and mass spectra. The physical data of all the newly reported derivatives were summarized in Table 1.

C. Molecular docking studies

Docking studies of synthesized pyran derivatives have been studied to find out the best drug moiety giving an insight into substituted and configurational needs for perfect receptor pit which cause the evolution of best pharmacophore moiety. These models are achieved to get more precise and consistent picture of the potentially active biological molecules at the atomic level and also produce new insights that could be used to develop new therapeutic agents.

In anticancer docking results, only best conformers were chosen and dock value for every ligand listed in Table 2. Most stable receptor ligand complex form is the one which has low dock value. The best docked conformer of every ligand and receptor were joined together and their complexes were optimized energetically after docking execution by define a radius of $10 \, \mathrm{A}^{\circ}$ measured from the docked ligand. Step by step optimization energy was done by using first hydrogen, side chains attached and finally the backbone of receptor molecule.

Fig. 2, shows the binding relationship between compound **4d** and protein kinase 1 active site residues interact with Arg 197, Thr 219, Asp117 and Ala 234.

TABLE II DOCKING SCORES OF THE SYNTHESIZED COMPOUNDS.

	Docking score (Binding energy)					
Compound	Breast Cancer	Colon Cancer	Lung Cancer	Pancreatic Cancer		
4a	-93.04	-86.24	-90.09	-83.87		
4b	-93.55	-92.00	-83.92	-83.54		
4c	-97.29	-94.78	-78.13	-78.34		
4d	-104.89	-89.55	-91.56	-86.26		

Important hydrogen bonding interaction takes place between Thr 219 and the (C=O) oxygen atom of pyran ring, another strong bonding takes place between Asp117 and Ala 234 and the nitrogen atom attached to the pyran ring.

III. CONCLUSIONS

In conclusion, it is found that a simple and efficient method for the synthesis of phenothiazine substituted different pyran derivatives under conventional heating and ultrasonic method. The main advantages of ultrasonic irradiation are those which include yield improvement, short reaction times, easy reaction set-up and use of very little amount of solvents or without solvents. The synthesized phenothiazine derivatives were subjected to anticancer molecular docking studies using four dissimilar cancer target proteins. The results of anticancer docking studies showed that the synthesized compound 4d possess significant binding energy with breast cancer when comparing with other compound.

REFERENCES

- [1] P. Gabriel, R. Sarmiento, G. Vitale, A. Javier, Y. Graciel, A. Moltrasioe and G. Moglioni, "Synthesis and antifungal activity of some substituted phenothiazines and related compounds," *Eur. J. Med. Chem*, vol. 46, 2011, pp. 101-105.
- [2] J. Raval and K.K. Desai, "Synthesis and antimicrobial activity of new triazolopyridinyl phenothiazines," *ARKIVOC*, vol. 2005, 2005, pp. 21-28.
- [3] A.R. Trivedi, A.B. Siddiqui and V.H. Shah, "Design, synthesis, characterization and antitubercular activity of some 2-heterocycle-substituted phenothiazines," *ARKIVOC*, vol. 2008, 2008, pp. 210-217.
- [4] S. Sharma, V.K. Srivastava and A. Kumar, "Synthesis and anti-inflammatory activity of some heterocyclic derivatives of phenothiazine," *Pharmazie*, vol. 60, 2005, pp. 18-22.
- [5] M. Kalkanidis, N. Kloni, L. Tilley, L.W. Deady, M. Kalkanidis, N. Klonis, L. Tilley and L.W. Deady, "Novel phenothiazine antimalarials: synthesis, antimalarial activity, and inhibition of the formation of beta-haematin," *Bio. Chem Pharmacol*, vol. 63, 2002, pp. 833-842.
- [6] G. Lin, K.K. Midha and E.M. Hawes, "Synthesis of the piperidinone metabolites of piperidine type phenothiazine antipsychotic drugs via ruthenium tetroxide oxidation," *J. Heterocycl. Chem*, vol. 28, 1991, pp. 215-219.
- [7] N. Tanaji, V. Jayant and G. Vaijanath, "Synthesis and antimicrobial activity of some new N-acyl substituted phenothiazines," *Eur. J. Med. Chem*, vol. 44, 2009, pp. 5094-5098.
- [8] M. Viveros and L. Amaral, "Enhancement of antibiotic activity against poly-drug resistant Mycobacterium tuberculosis by phenothiazines." *Int. J. Antimicrob. Ag*, vol. 17, 2001, pp. 225-228.
- [9] P.B. Madrida, W.E. Polgara, L. Toll and M.J. Tanga, "Synthesis and antitubercular activity of phenothiazines with reduced binding to dopamine and serotonin receptors," *Bioorg. & Med. Chem. Lett*, vol. 17, 2007, pp. 3014-3017.
- [10] P. Krystian, J. Magorzata, M. Beata, Z. Micha, A. Jolanta and K. Maja, "Anticancer activity of newly synthesized azaphenothiazines from NCI's anticancer screening bank" *Pharmacol. Rep.*, vol. 62, 2010, pp. 319-332.
- [11] S. Daversh, R.S. McDonald, A. Penwell, S. Conrad and K.V. Darvesh, "Structure-activity relationships for inhibition of

- human cholinesterases by alkyl amide phenothiazine derivatives," *Biorg. Med. Chem*, vol. 13, 2005, pp. 211-222.
- [12] M. Schmidt, M. Teitge, M.E. Castillo, T. Brandt, B. Dobner and A. Langner, "Synthesis and biochemical characterization of new phenothiazines and related drugs as MDR reversal agents," *Arch. Pharm. (Weinheim)*, vol. 341, 2008, pp. 624-638.
- [13] A. Bissi, M. Meli, S. Gobbi, A. Rampa, M. Tolomeo and L. Dusonchet, "Multidrug resistance reverting activity and antitumor profile of new phenothiazine derivatives," *Bioorg. Med. Chem*, vol. 16, 2008, pp. 6474-6482.
- [14] M.A. Cristina, B. Elena, R. Benoît, G. Philippe, B. Dalila, F. Amaury, D. Joelle and G. Alina, "Synthesis and anticancer activity of analogues of phenstatin, with a phenothiazine Aring, as a new class of microtubule-targeting agents," *Bioorg. & Med. Chem. Lett*, vol. 23, 2013, pp. 147-152.
- [15] I.K. Pajeva, M. Wiese, H.P. Cordes and J.K. Seydel, "Membrane interactions of some catamphiphilic drugs and relation to their multidrug-resistance-reversing ability," *J. Cancer. Res. Clin. Oncol*, vol. 122, 1996, pp. 27-40.
- [16] A. Ramu and N. Ramu, "Reversal of multidrug resistance by phenothiazines and structurally related compounds," *Cancer Chemother. Pharmacol*, vol. 30, 1992, pp. 165-173.
- [17] K. Pluta, M. Jeleñ, B. Morak Modawska, M. Zimecki, J. Artym and M. Kocieba, "Anticancer activity of newly synthesized azaphenothiazines from NCI's anticancer screening bank," *Pharmacol Rep*, vol. 62, 2010, pp. 319-332.
- [18] A. Seelig, R. Gottschlich and R.M. Devant, "A method to determine the ability of drugs to diffuse through the bloodbrain barrier," *Proc. Natl. Acad. Sci. USA*, vol. 91, 1994, pp. 68-72.
- [19] K.H. Chen, C.E. Lin, W.S. Liao, W.Y. Lin and Y.Y. Hsiao, "Separation and migration behavior of structurally related phenothiazines in cyclodextrin-modified capillary zone electrophoresis," *J. Chromatogr. A*, vol. 979, 2002, pp. 399-408.
- [20] L. Danielson and M.A. Lill, "New computational method for prediction of interacting protein loop regions," *Proteins: Struct. Funct. Bioinf*, vol. 78, 2010, pp. 1748-1759.
- [21] M.A. Lill and M.L. Danielson, "Computer-aided drug design platform using PyMOL," J. Com. Aid. Mol. Des, vol. 25, 2011, pp. 13-19.